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Mucosal Leishmaniasis Caused by Leishmania (Viannia) braziliensis and Leishmania (Viannia) guyanensis in the Brazilian Amazon

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Abstract

Background: Leishmania (Viannia) braziliensis is a parasite recognized as the most important etiologic agent of mucosal leishmaniasis (ML) in the New World. In Amazonia, seven different species of Leishmania, etiologic agents of human Cutaneous Leishmaniasis, have been described. Isolated cases of ML have been described for several different species of Leishmania: L. (V.) panamensis, L. (V.) guyanensis and L. (L.) amazonensis.

Methodology: Leishmania species were characterized by polymerase chain reaction (PCR) of tissues taken from mucosal biopsies of Amazonian patients who were diagnosed with ML and treated at the Tropical Medicine Foundation of Amazonas (FMTAM) in Manaus, Amazonas state, Brazil. Samples were obtained retrospectively from the pathology laboratory and prospectively from patients attending the aforementioned tertiary care unit.

Results: This study reports 46 cases of ML along with their geographical origin, 30 cases caused by L. (V.) braziliensis and 16 cases by L. (V.) guyanensis. This is the first record of ML cases in 16 different municipalities in the state of Amazonas and of simultaneous detection of both species in 4 municipalities of this state. It is also the first report of ML caused by L. (V.) guyanensis in the states of Pará, Acre, and Rondônia and cases of ML caused by L. (V.) braziliensis in the state of Rondônia.

Conclusions/Significance: L. (V.) braziliensis is the predominant species that causes ML in the Amazon region. However, contrary to previous studies, L. (V.) guyanensis is also a significant causative agent of ML within the region. The clinical and epidemiological expression of ML in the Manaus region is similar to the rest of the country, although the majority of ML cases are found south of the Amazon River.


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Introduction

Mucosal leishmaniasis (ML) in the Americas is mainly associated with L. (V.) braziliensis, the species recognized as the most important etiologic agent of the disease [1,2]. Marzochi and Marzochi [3], based on the epidemiological and geographical distribution of that same species in different ecosystems, suggested that the human disease emerged in the Western Amazon, in particular south of the Amazon River, where L. (V.) braziliensis is the predominant form. Here the majority of patients with ML typically work in areas of primary rainforest, involved in activities related to forest product extraction [4,5]; in these cases, the mucosal disease is the outcome of patients with a history of skin lesions that were not treated properly. Because of this, ML is an important public health problem and neglected disease in the Brazilian Amazon [5,6]. L. (V.) panamensis, L. (V.) guyanensis and L. (L.) amazonensis have also been associated with ML, but very few cases of ML have been associated with L. (V.) guyanensis [7,8,9]. Early diagnosis and access to treatment of cutaneous leishmaniasis (CL) are crucial to avoid the development of ML and complications of this form of the disease, given its complexity and severity. In an attempt to improve diagnosis, molecular techniques such as Polymerase Chain Reaction (PCR) have been developed for the detection of Leishmania parasites in clinical samples [10,11]; however, the low amount of DNA found in paraffin tissue hinders the characterization of species [12]. The identification of parasite species, today most commonly from genetic analyses, can directly contribute to our understanding of the epidemiology of leishmaniasis [13,14,15,16]. The aim of this study is to describe the distribution of Leishmania species in Amazonian patients with ML.
that were treated at the Tropical Medicine Foundation of Amazonas (FMTAM), a tertiary care unit, while taking into consideration the geographical origin of each case.

Materials and Methods

Ethics Statement

This study was prepared in accordance with international ethical guidelines for biomedical research involving human subjects. The project was approved for retrospective and prospective study; retrospective study in July 1992 to June 2006 and prospective study from July 2006 to December 2008. For the retrospective study, the samples (paraffin biopsies) were obtained from an already-existing collection in the pathology laboratory of FMTAM. For the prospective study, samples were obtained from patients presenting to FMTAM following informed consent, which was documented and signed.

Study Design

The study population consists of patients with ML who were diagnosed and treated at the FMTAM in the city of Manaus, Amazonas state, Brazil, from July 1992 to December 2006. All patients came from the Brazilian Amazon. This region covers an area of 5,000,000 km², 59% of Brazil’s territory, and contains over 775 municipalities in the states of Amazonas, Amapá, Mato Grosso, Western Maranhão, Pará, Rondônia, Roraima, Acre, and Tocantins. The total population for the region has been estimated at 20.3 million people – 68.9% of whom reside in urban areas while the remaining 31.1% reside in rural areas.

The distribution of cases was initially based on the municipality where patients with a prior history of CL acquired their cutaneous lesions that subsequently developed into mucosal disease. In patients with no prior history of CL, the following exposure factors were considered to be more important than place of birth – living within an endemic area and a history of exposure factor activities in natural resource extraction in areas of natural forest.

DNA Preparation

The biopsied tissues were preserved in three different media: a) formalin-fixed paraffin-embedded, b) imprint tissue on filter paper, or c) in buffer L6 [18]. The methodology for the extraction of DNA varied according to preservation methodology.

Embedded in paraffin. We performed 12 cuts of 20 μm in each block of embedded tissue using a disposable blade for each block. The samples were deposited in 1.5 mL Eppendorf tubes. The deparaffinization was done with xylol and the DNA extraction using the protocol of the Dneasy Blood & Tissue Kit (Qiagen).

Filter paper. The material from the filter paper was cut and placed in sterile Eppendorf tubes. DNA was extracted using the “blood spot” protocol of the PureLink Genomic DNA kit (Invitrogen).

Biopsies solution L6. Excess solution was removed by centrifugation at 14,000 rpm. The supernatant was discarded and the tissue was homogenized using individual disposable test tubes (Anachem). The tissue was then processed using the DNeasy Blood & Tissue Kit (Qiagen) as described by the manufacturer.

DNA Amplification by PCR

The presence of Leishmania DNA in tissue samples was detected by PCR using genus-specific primer 13a and 13b [19] according to the protocol described by Reale et al. [20].

In all tissue samples that were positive for Leishmania, PCR-RFLP was used to identify each species present in the biopsy. PCR was performed as described by Marfurt et al. [21]. DNA was amplified using primers Fme and Rme. Ten μl of the PCR products were digested with 1 U HaelI and 1 U NcoI (New England Biolabs) at 37°C for 2 hours and 30 minutes. The resulting restriction fragments were separated on a 2.5% agarose gel. The size of the fragments was estimated by comparison with a 100 bp DNA ladder and compared with positive controls for L. (V.) braziliensis and L. (V.) guyanensis.

As positive controls for DNA extraction, all DNA samples that did not amplify Leishmania-PCR using 13a and 13b primers were subjected to PCR targeting 147 bp fragments of human actin gene. The sequence of primer used was Hu_actin1_fwd 5'-CTGTTGGCATCCAGAACTA-3’ and Hu_actin1_rev 5'-AGGGCAGTGATCTCCTTCTG-3’. The PCR reaction was performed in a volume of 25μl, 18.75 μl H2O, 2.50 μl 10x buffer containing each primer 0.3 μl, 3.5μl MgCl2, 0.2 mM dNTPs and 1 U Platinum Taq DNA Polymerase (Invitrogen) 2 μl DNA template. The PCR conditions were 5 minutes at 94°C followed by 40 cycles of 35 seconds at 94°C, 30 seconds at 58°C and 30 seconds at 70°C, and a final extension at 70°C, 7 minutes.

Results

The reported 46 cases of ML caused by L. (V.) guyanensis and L. (V.) braziliensis, along with their geographical origin, are depicted in Figure 1. This is the first record of ML cases in 16 different municipalities in the state of Amazonas and of simultaneous detection of both species in 4 municipalities of this state. It is also the first record of ML caused by L. (V.) guyanensis in the states of Pará, Acre, and Rondônia and cases of ML caused by L. (V.) braziliensis in the state of Rondônia. Thirty eight patients had a previous history of CL. Thirteen patients were from municipalities located north of the Amazon River and 33 patients came from south of the river.

Comparing the two characterized species revealed no differences concerning clinical and epidemiological aspects of cases studied (Table 1).
Within the group of patients studied, 39 had a history of previous CL and the source of origin of the mucosal disease was considered to be the same municipality where the cutaneous form was acquired. Of the seven patients without a previous history of CL, six lived and worked their whole lives in the same place and only went to Manaus for treatment; the remaining patient lived in a rural area of Manaus.

Among the 46 patients, 38 were male and 8 were female. The average period mediating skin to mucosal disease was 17.9 years (range: 4 months to 74 years) and the average duration of the mucosal disease was 8.3 years (30 days to 39 years). Two of the female patients had concomitant disease (CL/ML). Both were pregnant at the time of acquiring CL and therefore did not treat their skin lesions. The average age of the study population was 47.5 years (range: 16 to 80).

The PCR, performed on samples of 143 patients, was positive in 56 individuals, but in 10 samples it was not possible to characterize the species, probably due to the low amount of DNA or a consequence of formalin fixation and paraffin embedded tissues. Nine patients received adequate treatment of their CL and therefore did not treat their skin lesions. The average age of the study population was 47.5 years (range: 16 to 80).

The PCR, performed on samples of 143 patients, was positive in 56 individuals, but in 10 samples it was not possible to characterize the species, probably due to the low amount of DNA or a consequence of formalin fixation and paraffin embedded tissues. Nine patients received adequate treatment of their CL and therefore did not treat their skin lesions. The average age of the study population was 47.5 years (range: 16 to 80).

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Discussion

Leishmaniasis is a disease that is increasing in the Northern Hemisphere as a result of tourism and armed conflict in tropical regions [22,23]. Cases of ML have been associated with multiple species [7,8,9], but our record of cases of ML caused by L. (V.) guyanensis is unusual. It was previously believed that the occurrence of ML caused by L. (V.) guyanensis was extremely low, with only isolated cases having been described. The past low infection rate described in the literature is likely to be the result of limited studies on ML in the regions where L. (V.) guyanensis is endemic.

The geographic distribution of cases of American tegumentary leishmaniasis indicates that L. (V.) braziliensis is the predominant species south of the Amazon River [24], while studies in the
Manaus region (north of the Amazon River) [18,25] found that \textit{L. (V.) guyanensis} is the most common species. In this study, 32 (71.1\%) of the ML cases are from the south of the river: 22 (68.7\%) were caused by \textit{L. (V.) braziliensis} and 10 (31.3\%) by \textit{L. (V.) guyanensis}. North of the Amazon River, 13 (28.9\%) patients were infected: 8 (61.5\%) with \textit{L. (V.) braziliensis} and 5 (38.5\%) with \textit{L. (V.) guyanensis} (Figure 1), which were mainly found in the Manaus area. This data suggests that no major differences exist between north and south of the river regarding the distribution of species causing ML.

The association between mucosal disease and previous skin lesions is widely accepted, as both forms can be caused by a single species [26,27], and indeed in the 46 cases described here 37 had a previous history of CL. In the eastern Brazilian Amazon, mucosal disease occurs in patients with a history of previous skin lesions that were either untreated or treated inappropriately, and which were often caused by \textit{L. (V.) braziliensis} [2,26]. The data from this study on patient age, and the relationship between a previous history of CL and ML, are in support of previous findings [9,28].

The current study extends this information and contributes new data on the distribution of \textit{L. (V.) braziliensis} in western Amazonia, providing the first record of this species in 16 municipalities of Amazonas state and an additional 12 municipalities in three other states in the region: Acre – 1 case, Rondônia – 5 cases and Pará – 7 cases. It is also very important to emphasize the record of 16 ML cases caused by \textit{L. (V.) guyanensis} in six different municipalities in Amazonas state, three in Pará, one in Rondônia and one in Acre (Figure 1).

It is probable that ML caused by \textit{L. (V.) guyanensis} has always existed in Amazonia. We believe that this study fills a gap in knowledge about the epidemiology of ML, rather than identifying a change in disease pattern. Although this work has not assessed the genetic polymorphism of \textit{L. (V.) guyanensis}, this has already been demonstrated [29,30] and others have demonstrated this with respect to \textit{L. (V.) braziliensis} [29,31,32,33]. The finding of several hybrid genotypes of \textit{Leishmania (Viannia)} in foci of cutaneous and mucocutaneous leishmaniasis has also been reported [34]. One cannot exclude the possibility of a genetic polymorphism of \textit{L. (V.) guyanensis} in the etiology of ML in the Amazonian region, since little has been reported prior to this study.

The association between inappropriately treated cutaneous forms of the disease and the occurrence of ML appears to be maintained for both \textit{L. (V.) guyanensis} and \textit{L. (V.) braziliensis}. The association between cutaneous forms treated inappropriately and the occurrence of the mucosal form, also in ML caused by \textit{L. (V.) guyanensis}, seems to keep the same relationship observed for the \textit{L. (V.) braziliensis}. However, it should be noted that poor access to the diagnosis and treatment of leishmaniasis is common in the Amazon region. This is due to the isolation of communities, with access being almost exclusively by boat in many areas. Furthermore, many patients lack the financial resources to stay for long periods in Manaus to ensure adequate treatment and follow-up. These factors may be associated with the development of mucosal disease. The high prevalence in males in our study population has also been observed by other authors [35,36]. The average time of 17.6 years between the diagnosis of CL and the appearance of ML (with one patient having a 74-yr gap between CL and ML) recorded in this study is also in agreement with previous findings [37,38] on the persistence of this parasite in the host’s body and the subsequent triggering of mucosal disease.

From a clinical point of view, we would like to draw the reader’s attention to the large number of cases – 21 (45.7\%) – with nasal perforation, which supports previous findings [27] that have demonstrated the potential of this species to cause more severe disease.

In summary, based on the results of this study, \textit{L. (V.) braziliensis}, which caused 2/3 of the studied cases, is the predominant species

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**Table 1. Clinical and epidemiological aspects of cases according to the species identified.**

<table>
<thead>
<tr>
<th>Category</th>
<th>Subcategory</th>
<th>\textit{L. (V.) braziliensis}</th>
<th>\textit{L. (V.) guyanensis}</th>
</tr>
</thead>
<tbody>
<tr>
<td>History of CL</td>
<td>Yes</td>
<td>24 (80%)</td>
<td>15 (93.8%)</td>
</tr>
<tr>
<td></td>
<td>No</td>
<td>6 (20%)</td>
<td>1 (6.2%)</td>
</tr>
<tr>
<td>Location of lesions</td>
<td>Nasal</td>
<td>26 (86.7%)</td>
<td>11 (68.8%)</td>
</tr>
<tr>
<td></td>
<td>Nasal/ oropharynx</td>
<td>3 (10%)</td>
<td>2 (12.5%)</td>
</tr>
<tr>
<td></td>
<td>Nasal/ pharynx / larynx</td>
<td>1 (3.3%)</td>
<td>2 (12.5%)</td>
</tr>
<tr>
<td></td>
<td>Oropharynx</td>
<td>0 (0%)</td>
<td>1 (6.2%)</td>
</tr>
<tr>
<td>Clinical presentation</td>
<td>Infiltrated</td>
<td>3 (10%)</td>
<td>3 (18.8%)</td>
</tr>
<tr>
<td></td>
<td>Perforation</td>
<td>14 (46.7%)</td>
<td>6 (37.5%)</td>
</tr>
<tr>
<td></td>
<td>Ulcer</td>
<td>13 (43.3%)</td>
<td>7 (43.8%)</td>
</tr>
<tr>
<td></td>
<td>Positive</td>
<td>1 (3.3%)</td>
<td>2 (12.5%)</td>
</tr>
<tr>
<td>Histop. examination</td>
<td>Compatible with ML</td>
<td>23 (7.7%)</td>
<td>8 (62.5%)</td>
</tr>
<tr>
<td></td>
<td>Inconclusive</td>
<td>1 (33.3%)</td>
<td>2 (12.5%)</td>
</tr>
<tr>
<td></td>
<td>Not performed</td>
<td>2 (6.7%)</td>
<td>2 (12.5%)</td>
</tr>
<tr>
<td></td>
<td>Non-specific chronic rhinitis</td>
<td>3 (10%)</td>
<td>2 (12.5%)</td>
</tr>
<tr>
<td>Treatment of CL</td>
<td>Regular</td>
<td>4 (16.7%)</td>
<td>5 (31.3%)</td>
</tr>
<tr>
<td></td>
<td>Irregular</td>
<td>20 (83.3%)</td>
<td>10 (62.5%)</td>
</tr>
<tr>
<td>Time between CL/ML</td>
<td>Median time (years)</td>
<td>16.9</td>
<td>14.9</td>
</tr>
<tr>
<td>Disease duration</td>
<td>Median time (years)</td>
<td>12</td>
<td>8.6</td>
</tr>
</tbody>
</table>

that causes ML in the Amazon region. However, contrary to previous studies, L. (V.) guyanensis is also a significant causative agent of ML in the region. The clinical and epidemiological expression of ML in the Amazon region is similar to the rest of the country, although the majority of ML cases are found south of the Amazon River. ML infections are much more common in men than in women, and men also tend to develop more severe forms of disease with a high incidence of perforation and involvement of structures outside of the nasal cavity.

Supporting Information

Checklist S1 Struev Checklist

Found at: doi:10.1371/journal.pntd.0000980.s001 (0.06 MB DOC)

Author Contributions

Conceived and designed the experiments: JAdOG LIdARCC LGdLF. Performed the experiments: JAdOG SRP HS LGdLF. Analyzed the data: JAdOG PG AM VA MdGVB LGdLF. Contributed reagents/materials/analysis tools: JAdOG MdGVB LGdLF. Wrote the paper: JAdOG MdGVB LGdLF.

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